

December 28, 2020

Prince George's County Public School
Environmental Safety Office
13306 Old Marlboro Pike
Upper Marlboro, MD 20772

Attention: Alex Baylor
alex.baylor@pgcps.org

Subject: Indoor Air Quality Survey
Scotchtown Hills Elementary School
15950 Dorset Road
Laurel, MD 20707

Mr. Baylor:

On December 8, 2020, a Soil and Land Use Technology, Inc. (SaLUT) Industrial Hygienist conducted an indoor air quality (IAQ) evaluation at Scotchtown Hills Elementary School, a property maintained by Prince George's County Public School (PGCPS) located at 15950 Dorset Road, Laurel, MD 20707. The inspection was performed in accordance with PGCPS contract number IFB 022-19.

Methodology

The IAQ evaluation conducted by SaLUT included a visual assessment, IAQ instrumentation screening, and a collection of interior air samples for mold in representative locations throughout the building. Additionally, one building exterior environmental air sample was taken for comparison.

Air-borne fungal spore samples were collected on *Air-O-Cell* cassettes using a Buck BioAire calibrated pump. The air samples were taken between three and five feet from the ground. In tandem with collecting mold samples, real-time readings for carbon dioxide, carbon monoxide, temperature and relative humidity were collected using a Fluke 975 Air Meter in representative areas within the facility.

The fungal spore air samples were delivered to EMSL Analytical, Inc. of Beltsville, Maryland for analysis. Fungal spores and particulates in air samples were analyzed by Optical Microscopy (methods EMSL 05-TP-003 and ASTM D7391). The sample chain-of-custody and laboratory reports are attached.

Observations

The table below summarizes the main observations from the IAQ survey at Scotchtown Hills Elementary School, visited on December 8, 2020.

Table 1-Observations

Location	Summary of Observations 12-08-2020
Cafeteria	2'x4' ceiling tiles and 1'x1' tile floor; No visual signs of microbial growth; Mild odor; Stained ceiling tiles; No visible dust on floor/other furniture surfaces; No visible dust around ventilator; Central AC.
Hallway next to Media Center room	2'x4' ceiling tiles and terrazzo floor; No visual signs of microbial growth, and no odor; No visible dust on floor/other furniture surfaces; No visible dust around ventilator; Central AC.
Hallway next to Classroom 3	2'x4' ceiling tiles and terrazzo Floor; No visual signs of microbial growth, and no odor; No visible dust on floor/other furniture surfaces; No visible dust around ventilator; Central AC.
2 nd floor, Hallway next to Classroom 15	2'x4' ceiling tiles and 1'x 1' tile floor; No visual signs of microbial growth, and no odor; No visible dust on floor/other furniture surfaces; No visible dust around ventilator; Central AC.
2 nd floor, Hallway next to Classroom 28	2'x4' ceiling tiles and 1'x 1' tile floor; No visual signs of microbial growth, and no odor; No visible dust on floor/other furniture surfaces; No visible dust around ventilator; Central AC.

Measurements of Indoor Environmental Quality Parameters

Table 2 depicts a summary of average measurements of comfort.

Temperature

The American Society of Heating, Refrigerating and Air-Conditioning Engineers (ASHRAE) have published recommendations for year round acceptable temperatures in Standard 55-2010 *Thermal Environmental Conditions for Human Occupancy*. The winter comfort range is 20 to 24°C (68 to 75°F) and 23 to 26°C (73 to 79°F) is the summer comfort range. The temperature readings were lower than the ASHRAE recommended ranges in the representative spaces.

Relative Humidity (RH)

RH is a key factor for mold growth. Mold has the potential of growing on suitable surfaces with humidity levels above 60%. ASHRAE Standard 62.1-2010 *Ventilation for Acceptable Indoor Air Quality* recommends a maximum indoor RH of 65% to preclude the likelihood of condensation on cool surfaces encouraging mold growth. The RH readings were within the ASHRAE recommended ranges in the representative areas.

Carbon Dioxide (CO₂)

Under conditions of maximum occupancy, ASHRAE Standard 62.1-2010, Appendix C, infers that the acceptable CO₂ upper limit is the prevailing outdoor CO₂ concentration plus 700 parts per million (ppm). On the day of the space evaluation, the outdoor (building exterior) CO₂ concentration was approximately 480 ppm therefore indoor concentrations should not exceed approximately 1,180 ppm (700 + 480). The maximum average interior CO₂ concentration detected was 472 ppm in on the 2nd floor in the Hallway next to Classroom 15 and in the Hallway next to Classroom 28, a range within the ASHRAE recommendations, per Table 2 below.

Carbon Monoxide (CO)

CO is a colorless and odorless gas that is produced by the incomplete combustion of carbon containing fuels. Oil, gasoline, diesel fuels, wood, coke, and coal are major sources of CO. All registered CO concentrations were below the EPA National Ambient Air Quality Standard (NAAQS) of 9 ppm, per Table 2 below.

**Table 2: Scotchtown Hills Elementary School, Instrumental Screening Levels
December 8, 2020 (7:30 AM-9:30 AM)**

Sample Location	Temp °F	RH%	CO ppm	CO ₂ ppm
Standards	ASHRAE 68 to 75°F*	ASHRAE <65%	NAAQS 9	ASHRAE 1,180
Cafeteria	59.9	37.9	0	465
Hallway next to Media Center room	59.0	38.0	0	470
Hallway next to Classroom 3	58.7	37.5	0	468
2nd floor, Hallway next to Classroom 15	59.8	38.0	0	472
2nd floor, Hallway next to Classroom 28	60.0	38.0	0	472
Outside Exterior EV sample	56.2	34.0	0	480

PM - Particulate Matter size
°F - Degrees Fahrenheit
CO - Carbon Monoxide
ppm - parts per million

µg/m³ - micrograms per cubic meter
RH% - % Relative Humidity
CO₂ - Carbon Dioxide
* - Winter Comfort Range

Mold-in-Air Samples

There are no definitive regulations or standardized guidelines for addressing airborne mold in an indoor setting. If building systems (ventilation, envelope) are functioning properly, the indoor population profile should mimic what is encountered outdoors and the concentrations should be below the outdoor (building exterior) environmental sample levels.

Table 3 summarizes airborne mold spore sampling results and locations. On December 8, 2020, total mold counts in representative samples (spore count/m³ of air) in all the areas inspected were lower than the outdoor concentrations. Laboratory analysis follows this report (see attachment).

**Table 3: Scotchtown Hills Elementary School - Measurements of Mold-in-Air Samples
December 8, 2020 (7:30 AM-9:30 AM)**

Spore Types	Cafeteria	Hallway next to Media Center	Hallway next to Classroom 3	Hallway next to Classroom 15
<i>Alternaria (Ulocladium)</i>	-	-	-	-
<i>Ascospores</i>	-	-	-	-
<i>Aspergillus/Penicillium</i>	40	-	90	-
<i>Basidiospores</i>	40	40	-	-
<i>Bipolaris++</i>	-	-	-	-
<i>Chaetomium</i>	-	-	-	-
<i>Cladosporium</i>	-	-	40	-
<i>Curvularia</i>	-	-	-	-
<i>Epicoccum</i>	-	-	-	-
<i>Fusarium</i>	-	-	-	-
<i>Ganoderma</i>	-	-	-	-
<i>Myxomycetes++</i>	-	-	-	-
<i>Pithomyces++</i>	-	-	-	-
<i>Rust</i>	-	-	-	-
<i>Scopulariopsis/Microascus</i>	-	-	-	-
<i>Stachybotrys/Memnoniella</i>	30	-	-	-
<i>Unidentifiable Spores</i>	-	-	-	-
<i>Zygomycetes</i>	-	-	-	-
<i>Nigrospora</i>	-	-	-	-
<i>Hyphal Fragment</i>	-	-	-	-
<i>Insect Fragment</i>	40	-	-	-
<i>Pollen</i>	-	-	-	-
Total Fungi	110	40	130	None Detect

* Spore Counts per cubic meter of air (Counts/m³).

++Includes other spores with similar morphology.

**Table 3: Scotchtown Hills Elementary School -
Measurements of Mold-in-Air Samples continued
December 8, 2020 (7:30 AM-9:30 AM)**

Spore Types	Hallway next to Classroom 28	Outside EXT EV sample	Field Blank
<i>Alternaria (Ulocladium)</i>	-	10	-
<i>Ascospores</i>	-	300	--
<i>Aspergillus/Penicillium</i>	-	-	-
<i>Basidiospores</i>	-	90	-
<i>Bipolaris++</i>	-	-	-
<i>Chaetomium</i>	-	-	-
<i>Cladosporium</i>	-	300	-
<i>Curvularia</i>	-	-	-
<i>Epicoccum</i>	-	-	-
<i>Fusarium</i>	-	-	-
<i>Ganoderma</i>	-	-	-
<i>Myxomycetes++</i>	-	-	-
<i>Pithomyces++</i>	-	40	-
<i>Rust</i>	-	90	-
<i>Scopulariopsis/Microascus</i>	-	-	-
<i>Stachybotrys/Memnoniella</i>	-	-	-
<i>Unidentifiable Spores</i>	-	-	-
<i>Zygomycetes</i>	-	-	-
<i>Nigrospora</i>	-	-	-
<i>Hyphal Fragment</i>	-	40	-
<i>Insect Fragment</i>	-	-	-
<i>Pollen</i>	-	-	-
Total Fungi	None Detect	1480	No Trace

*Spore Counts per cubic meter of air (Counts/m³).

++Includes other spores with similar morphology.

Findings and Conclusions

The comfort parameters (i.e., temperature, RH, CO₂, and CO levels) in the representative areas conform to ASHRAE and/or NAAQS guidelines. On December 8,, 2020, total mold counts in representative area samples (spore count/m³ of air) in all the areas inspected were lower than the outdoor concentrations, indicating no amplified mold growth.

Thank you for the opportunity to provide industrial hygiene services for PGCPs. If you have any questions, please contact me at 301.595.3783.

Sincerely,



Chaminda Jayatilake, PE, CIH, CSP, CHMM
Certified Industrial Hygienist
Soil and Land Use Technology Inc. (SaLUT)

Attachment

Attachment - Mold Spore Sample Analytical Results and Chain-of-Custody Forms

Attachment

Mold Spore Sample Analytical Results and Chain-of-Custody Forms



EMSL Analytical, Inc.

10768 Baltimore Avenue Beltsville, MD 20705

Tel/Fax: (301) 937-5700 / (301) 937-5701

<http://www.EMSL.com> / beltsvillelab@emsl.com

EMSL Order: 192012075

Customer ID: SALU50

Customer PO:

Project ID:

Attention: Indika Jayatilake

SaLUT

1818 New York Avenue, NE

Suite 231

Washington, DC 20002

Project: Scotchtown Hills ES/ PGCPs IAQ 15950 Dorset Rd, Laurel, MD 20707

Phone: (301) 595-3783

Fax: (301) 595-3787

Collected Date: 12/08/2020

Received Date: 12/08/2020 02:50 PM

Analyzed Date: 12/10/2020

Test Report: Air-O-Cell™ Analysis of Fungal Spores & Particulates by Optical Microscopy (Methods MICRO-SOP-201, ASTM D7391)

Lab Sample Number:	192012075-0001			192012075-0002			192012075-0003		
Client Sample ID:	01			02			03		
Volume (L):	75			75			75		
Sample Location:	Cafeteria			H/W next to Media center room			H/W next to room 3		
Spore Types	Raw Count	Count/M ³	% of Total	Raw Count	Count/M ³	% of Total	Raw Count	Count/M ³	% of Total
Alternaria (Ulocladium)	-	-	-	-	-	-	-	-	-
Ascospores	-	-	-	-	-	-	-	-	-
Aspergillus/Penicillium	1	40	36.4	-	-	-	2	90	69.2
Basidiospores	1	40	36.4	1	40	100	-	-	-
Bipolaris++	-	-	-	-	-	-	-	-	-
Chaetomium	-	-	-	-	-	-	-	-	-
Cladosporium	-	-	-	-	-	-	1	40	30.8
Curvularia	-	-	-	-	-	-	-	-	-
Epicoccum	-	-	-	-	-	-	-	-	-
Fusarium	-	-	-	-	-	-	-	-	-
Ganoderma	-	-	-	-	-	-	-	-	-
Myxomycetes++	-	-	-	-	-	-	-	-	-
Pithomyces++	-	-	-	-	-	-	-	-	-
Rust	-	-	-	-	-	-	-	-	-
Scopulariopsis/Microascus	-	-	-	-	-	-	-	-	-
Stachybotrys/Memnoniella	2*	30*	27.3	-	-	-	-	-	-
Unidentifiable Spores	-	-	-	-	-	-	-	-	-
Zygomycetes	-	-	-	-	-	-	-	-	-
Arthrinium	-	-	-	-	-	-	-	-	-
Monodictys	-	-	-	-	-	-	-	-	-
Total Fungi	4	110	100	1	40	100	3	130	100
Hyphal Fragment	-	-	-	-	-	-	-	-	-
Insect Fragment	1	40	-	-	-	-	-	-	-
Pollen	-	-	-	-	-	-	-	-	-
Analyt. Sensitivity 600x	-	44	-	-	44	-	-	44	-
Analyt. Sensitivity 300x	-	13*	-	-	13*	-	-	13*	-
Skin Fragments (1-4)	-	1	-	-	1	-	-	1	-
Fibrous Particulate (1-4)	-	1	-	-	1	-	-	1	-
Background (1-5)	-	1	-	-	1	-	-	1	-

++ Includes other spores with similar morphology; see EMSL's fungal glossary for each specific category.

Abubakar Barry, Microbiology Laboratory Manager
or other Approved Signatory

No discernable field blank was submitted with this group of samples.

EMSL maintains liability limited to cost of analysis. Interpretation and use of test results are the responsibility of the client. This report relates only to the samples reported above, and may not be reproduced, except in full, without written approval by EMSL. EMSL bears no responsibility for sample collection activities or analytical method limitations. The report reflects the samples as received. Results are generated from the field sampling data (sampling volumes and areas, locations, etc.) provided by the client on the Chain of Custody. Samples are within quality control criteria and met method specifications unless otherwise noted. High levels of background particulate can obscure spores and other particulates, leading to underestimation. Background levels of 5 indicate an overloading of background particulates, prohibiting accurate detection and quantification. Present = Spores detected on overloaded samples. Results are not blank corrected unless otherwise noted. The detection limit is equal to one fungal spore, structure, pollen, fiber particle or insect fragment. "*" Denotes particles found at 300X. "-" Denotes not detected. Due to method stopping rules, raw counts in excess of 100 are extrapolated based on the percentage analyzed.

Samples analyzed by EMSL Analytical, Inc. Beltsville, MD AIHA-LAP, LLC-EMLAP Accredited #102891

Initial report from: 12/10/2020 04:55 PM

For information on the fungi listed in this report, please visit the Resources section at www.emsl.com



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Test Report: Air-O-Cell™ Analysis of Fungal Spores & Particulates by Optical Microscopy (Methods MICRO-SOP-201, ASTM D7391)

Lab Sample Number: Client Sample ID: Volume (L): Sample Location:	192012075-0004			192012075-0005			192012075-0006		
	Raw Count	Count/M³	% of Total	Raw Count	Count/M³	% of Total	Raw Count	Count/M³	% of Total
04 75 2nd floor H/W next to CR 15				05 75 2nd floor H/W next to CR 28			06 75 Outside Exterior EV Sample		
Spore Types									
Alternaria (Ullocladium)	-	-	-	-	-	-	1*	10*	0.7
Ascospores	-	-	-	-	-	-	7	300	20.3
Aspergillus/Penicillium	-	-	-	-	-	-	-	-	-
Basidiospores	-	-	-	-	-	-	2	90	6.1
Bipolaris++	-	-	-	-	-	-	-	-	-
Chaetomium	-	-	-	-	-	-	-	-	-
Cladosporium	-	-	-	-	-	-	6	300	20.3
Curvularia	-	-	-	-	-	-	-	-	-
Epicoccum	-	-	-	-	-	-	-	-	-
Fusarium	-	-	-	-	-	-	-	-	-
Ganoderma	-	-	-	-	-	-	-	-	-
Myxomycetes++	-	-	-	-	-	-	-	-	-
Pithomyces++	-	-	-	-	-	-	1	40	2.7
Rust	-	-	-	-	-	-	2	90	6.1
Scopulariopsis/Microascus	-	-	-	-	-	-	-	-	-
Stachybotrys/Memnoniella	-	-	-	-	-	-	-	-	-
Unidentifiable Spores	-	-	-	-	-	-	-	-	-
Zygomycetes	-	-	-	-	-	-	-	-	-
Arthrinium	-	-	-	-	-	-	14	610	41.2
Monodictys	-	-	-	-	-	-	1	40	2.7
Total Fungi	-	None Detect	-	-	None Detect	-	34	1480	100
Hyphal Fragment	-	-	-	-	-	-	1	40	-
Insect Fragment	-	-	-	-	-	-	-	-	-
Pollen	-	-	-	-	-	-	-	-	-
Analyt. Sensitivity 600x	-	44	-	-	44	-	-	44	-
Analyt. Sensitivity 300x	-	13*	-	-	13*	-	-	13*	-
Skin Fragments (1-4)	-	1	-	-	1	-	-	1	-
Fibrous Particulate (1-4)	-	-	-	-	1	-	-	1	-
Background (1-5)	-	1	-	-	1	-	-	1	-

++ Includes other spores with similar morphology; see EMSL's fungal glossary for each specific category.

Abubakar Barry, Microbiology Laboratory Manager
or other Approved Signatory

No discernable field blank was submitted with this group of samples.

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Samples analyzed by EMSL Analytical, Inc. Beltsville, MD AIHA-LAP, LLC-EMLAP Accredited #102891

Initial report from: 12/10/2020 04:55 PM

For information on the fungi listed in this report, please visit the Resources section at www.emsl.com



EMSL ANALYTICAL, INC.
LABORATORY PRODUCTS TRAINING

Microbiology Chain of Custody

EMSL Order Number (Lab Use Only):

192012075

PHONE:

FAX:

Company Name: SaLUT Inc.				EMSL-Bill to: <input checked="" type="checkbox"/> Same <input type="checkbox"/> Different If Bill to is Different note instructions in Comments**			
Street: 1818 New York Ave NE Suite 231				Third Party Billing requires written authorization from third party			
City: Washington		State/Province: DC		Zip/Postal Code: 20002		Country: USA	
Report To (Name): Indika Jayatilake				Telephone #: 301-595-3783			
Email Address: ijayatilake@salutinc.com				Fax #:		Purchase Order:	
Project Number/Location: Scotchtown Hills ES/PGCPS IAQ				Please Provide Results: <input type="checkbox"/> Fax <input checked="" type="checkbox"/> Email			
Location Address: 15950 Dorset Rd, Laurel, MD 20707				Connecticut Samples: <input type="checkbox"/> Commercial <input type="checkbox"/> Residential			
*Analysis completed in accordance with EMSL's Terms and Conditions located in the Analytical Price Guide. TATs are subject to methodology requirements							
Sterile, Sodium Thiosulfate Preserved Bottle Used: <input type="checkbox"/> Biocide Used in Source (specify): <input type="checkbox"/>							
Public Water Supply Samples: <input type="checkbox"/> Note: All results may automatically be reported to DOH if required by state.							
Turnaround Time (TAT) Options * - Please Check							
<input type="checkbox"/> 3 Hour	<input type="checkbox"/> 6 Hour	<input type="checkbox"/> 24 Hour	<input type="checkbox"/> 48 Hour	<input checked="" type="checkbox"/> 72 Hour	<input type="checkbox"/> 96 Hour	<input type="checkbox"/> 1 Week	<input type="checkbox"/> 2 Week
Microbiology Test Codes							
M001 Air-O-Cell		M174 MoldSnap		M024 Pseudomonas aeruginosa (MFT*)		M115 Sewage Screen - Water (P/A***)	
M030 Micro 5		M032 Allergenco-D		M015 Heterotrophic Plate Count		M116 Sewage Screen - Water (MPN**)	
M041 Fungal Direct Examination				M017 Total Coliform & E. coli (Colilert P/A***)		M117 Sewage Screen - Swab (P/A***)	
M169 Pollen ID & Enumeration				M018 Total Coliform & E. coli (MFT*)		M013 Sewage Screen - Swab (MFT*)	
M280 Dust Characterization Level-1				M114 Total Coliform & E. coli Enumeration (Colilert MPN**)		M133 Methicillin-resistant Staph. aureus (MRSA)	
M281 Dust Characterization Level-2				M019 Fecal Coliform (MFT*)		M031 Rapid-growing non-TB Mycobacteria Detection & Enumeration	
M005 Viable Fungi- Air Samples (Genus ID & Count)				M020 Fecal Streptococcus (MFT*)		M014 Endotoxin Analysis	
M006 Viable Fungi- Air Samples (Includes Penicillium, Aspergillus, Cladosporium, Stachybotrys Species ID & Count)				M029 Enterococci (MFT*)		M044 Group Allergen (Cat, Dog, Cockroach, Dust Mite)	
M007 Culturable fungi - Surface Samples (Genus ID & Count)				M129 Enterococci (Enterolert P/A***)		Other See Analytical Price Guide	
M008 Culturable fungi - Surface Samples (Includes Penicillium, Aspergillus, Cladosporium, Stachybotrys Species ID & Count)				M180 Real Time qPCR-ERMI 36 Panel		Legionella Analysis Please use EMSL Legionella COC	
M009 Bacteria Culture Gram Stain & Count				M025 Sewage Screen -Water (MFT*)			
M010 Bacteria Count & ID - 3 Most Prominent				*MFT= Membrane Filtration Technique			
M011 Bacteria Count & ID - 5 Most Prominent				**MPN= Most Probable Number			
M012 Pseudomonas aeruginosa (P/A***)				***P/A= Presence/Absence			
Name of Sampler: Jude Fonseka				Signature of Sampler:			
Sample #	Sample Location/Description	Sample Type	Potable/NonPotable (only for waters)	Test Code	Volume/Area	Date/Time Collected	Temperature (°C) (Lab Use Only)
01	Cafeteria	Air		M001	75L	12/8/2020	
02	H/W next to Media center room	Air		M001	75L	12/8/2020	
03	H/W next to room 3	Air		M001	75L	12/8/2020	
04	2nd floor H/W next to CR 15	Air		M001	75L	12/8/2020	
05	2nd floor H/W next to CR 28	Air		M001	75L	12/8/2020	
06	Outside Exterior EV Sample	Air		M001	75L	12/8/2020	
Client Sample # (s): -		Total # of Samples: 07		Samples Received Chilled? Yes / No			
Relinquished (Client):			Date:		Time:		
Received (Lab):			Date:		Time:		
Comments/Special Instructions:							

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 BELTSVILLE, MD
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