

December 28, 2020

Prince George's County Public School
Environmental Safety Office
13306 Old Marlboro Pike
Upper Marlboro, MD 20772

Attention: Alex Baylor
alex.baylor@pgcps.org

Subject: Indoor Air Quality Survey
Green Valley Administrative Offices
2215 Chadwick Street
Hillcrest Heights, MD 20748

Mr. Baylor:

On November 19, 2020, a Soil and Land Use Technology, Inc. (SaLUT) Industrial Hygienist conducted an indoor air quality (IAQ) evaluation at Green Valley Administrative Offices, a property maintained by Prince George's County Public School (PGCPS) located at 2215 Chadwick Street, Hillcrest Heights, MD 20748. The inspection was performed in accordance with PGCPS contract number IFB 022-19.

Methodology

The IAQ evaluation conducted by SaLUT included a visual assessment, IAQ instrumentation screening, and a collection of interior air samples for mold in representative locations throughout the building. Additionally, one building exterior environmental air sample was taken for comparison.

Air-borne fungal spore samples were collected on *Air-O-Cell* cassettes using a Buck BioAire calibrated pump. The air samples were taken between three and five feet from the ground. In tandem with collecting mold samples, real-time readings for carbon dioxide, carbon monoxide, temperature and relative humidity were collected using a Fluke 975 Air Meter in representative areas within the facility.

The fungal spore air samples were delivered to EMSL Analytical, Inc. of Beltsville, Maryland for analysis. Fungal spores and particulates in air samples were analyzed by Optical Microscopy (methods EMSL 05-TP-003 and ASTM D7391). The sample chain-of-custody and laboratory reports are attached.

Observations

The table below summarizes the main observations from the IAQ survey at , Green Valley Administrative Offices, visited on November 19, 2020.

Table 1-Observations

Location	Summary of Observations 11-19-2020
Cafeteria	2'x4' ceiling tiles and terrazzo floor; No visual signs of microbial growth, and mild odor; Stained ceiling tile; No visible dust on floor/other furniture surfaces; Central AC.
2 nd floor, Hallway next to Classroom 202	2'x4' ceiling tiles and terrazzo floor; No visual signs of microbial growth, and no odor; No visible dust on floor/other furniture surfaces; Central AC.
2 nd floor, Hallway next to CTE Storeroom 207	2'x4' ceiling tiles and terrazzo floor; No visual signs of microbial growth, and mild odor; No visible dust on floor/other furniture surfaces; Central AC.
Hallway next to Staff Lounge	2'x4' ceiling tiles and terrazzo floor; No visual signs of microbial growth, and mild odor; No visible dust on floor/other furniture surfaces; Central AC.
Hallway next to Classroom 404	2'x4' ceiling tiles and terrazzo floor; No visual signs of microbial growth, and mild odor; No visible dust on floor/other furniture surfaces; Central AC.

Measurements of Indoor Environmental Quality Parameters

Table 2 depicts a summary of average measurements of comfort.

Temperature

The American Society of Heating, Refrigerating and Air-Conditioning Engineers (ASHRAE) have published recommendations for year round acceptable temperatures in Standard 55-2010 *Thermal Environmental Conditions for Human Occupancy*. The winter comfort range is 20 to 24°C (68 to 75°F) and 23 to 26°C (73 to 79°F) is the summer comfort range. The temperature readings were within the ASHRAE recommended ranges in the representative spaces.

Relative Humidity (RH)

RH is a key factor for mold growth. Mold has the potential of growing on suitable surfaces with humidity levels above 60%. ASHRAE Standard 62.1-2010 *Ventilation for Acceptable Indoor Air Quality* recommends a maximum indoor RH of 65% to preclude the likelihood of condensation on cool surfaces encouraging mold growth. The RH readings were within the ASHRAE recommended ranges in the representative areas.

Carbon Dioxide (CO₂)

Under conditions of maximum occupancy, ASHRAE Standard 62.1-2010, Appendix C, infers that the acceptable CO₂ upper limit is the prevailing outdoor CO₂ concentration plus 700 parts per million (ppm). On the day of the space evaluation, the outdoor (building exterior) CO₂ concentration was approximately 416 ppm therefore indoor concentrations should not exceed approximately 1,116 ppm (700 + 416). The maximum average interior CO₂ concentration detected was 471 ppm in the Cafeteria, a range within the ASHRAE recommendations, per Table 2 below.

Carbon Monoxide (CO)

CO is a colorless and odorless gas that is produced by the incomplete combustion of carbon containing fuels. Oil, gasoline, diesel fuels, wood, coke, and coal are major sources of CO. All registered CO concentrations were below the EPA National Ambient Air Quality Standard (NAAQS) of 9 ppm, per Table 2 below.

**Table 2: Green Valley Administrative Offices - Instrumental Screening Levels
November 19, 2020 (7:30 AM-9:30 AM)**

Sample Location	Temp °F	RH%	CO ppm	CO ₂ ppm
Standards	ASHRAE 68 to 75°F*	ASHRAE <65%	NAAQS 9	ASHRAE 1,116
Cafeteria	69.8	29.2	0	471
2nd floor, Hallway next to Classroom 202	73.4	30.3	0	466
2nd floor, Hallway next to CTE Storeroom 207	71.6	31.7	0	436
Hallway next to Staff lounge	70.7	31.9	0	436
Hallway next to Classroom 404	68.9	42.7	0	434
Outside Exterior EV sample	55.4	43.7	0	416

PM - Particulate Matter size
°F - Degrees Fahrenheit
CO - Carbon Monoxide
ppm - parts per million

µg/m³ - micrograms per cubic meter
RH% - % Relative Humidity
CO₂ - Carbon Dioxide
* - Winter Comfort Range

Mold-in-Air Samples

There are no definitive regulations or standardized guidelines for addressing airborne mold in an indoor setting. If building systems (ventilation, envelope) are functioning properly, the indoor population profile should mimic what is encountered outdoors and the concentrations should be below the outdoor (building exterior) environmental sample levels.

Table 3 summarizes airborne mold spore sampling results and locations. On November 19, 2020, total mold counts in representative samples (spore count/m³ of air) in all the areas inspected were lower than the outdoor concentrations. Laboratory analysis follows this report (see attachment).

**Table 3: Green Valley Administrative Offices -
Measurements of Mold-in-Air Samples
November 19, 2020 (7:30 AM-9:30 AM)**

Spore Types	Cafeteria	2nd floor, Hallway next to Classroom 202	2nd floor, Hallway next to CTE Storeroom 207	Hallway next to Staff Lounge
<i>Alternaria (Ulocladium)</i>	-	-	-	-
<i>Ascospores</i>	-	-	-	-
<i>Aspergillus/Penicillium</i>	-	-	100	200
<i>Basidiospores</i>	100	300	100	100
<i>Bipolaris++</i>	-	-	-	-
<i>Chaetomium</i>	-	-	-	-
<i>Cladosporium</i>	100	40	100	40
<i>Curvularia</i>	-	-	-	-
<i>Epicoccum</i>	-	-	-	-
<i>Fusarium</i>	-	-	-	-
<i>Ganoderma</i>	-	-	40	-
<i>Myxomycetes++</i>	-	10	40	10
<i>Pithomyces++</i>	-	-	-	-
<i>Rust</i>	-	-	-	-
<i>Scopulariopsis/Microascus</i>	-	-	-	-
<i>Stachybotrys/Memnoniella</i>	-	-	-	-
<i>Unidentifiable Spores</i>	-	40	-	-
<i>Zygomycetes</i>	-	-	-	-
<i>Nigrospora</i>	-	-	-	-
<i>Hyphal Fragment</i>	-	80	-	-
<i>Insect Fragment</i>	-	-	10	-
<i>Pollen</i>	-	-	-	-
Total Fungi	200	390	390	670

* Spore Counts per cubic meter of air (Counts/m³).

++Includes other spores with similar morphology.

**Table 3: Green Administrative Valley Offices -
Measurements of Mold-in-Air Samples continued
November 19, 2020 (7:30 AM-9:30 AM)**

Spore Types	Hallway next to Classroom 404	Outside Exterior EV Sample	Field Blank
<i>Alternaria (Ulocladium)</i>	-	-	-
<i>Ascospores</i>	-	100	-
<i>Aspergillus/Penicillium</i>	680	300	-
<i>Basidiospores</i>	510	1100	-
<i>Bipolaris++</i>	-	40	-
<i>Chaetomium</i>	-	-	-
<i>Cladosporium</i>	80	930	-
<i>Curvularia</i>	-	-	-
<i>Epicoccum</i>	-	80	-
<i>Fusarium</i>	-	-	-
<i>Ganoderma</i>	-	-	-
<i>Myxomycetes++</i>	80	680	-
<i>Pithomyces++</i>	40	-	-
<i>Rust</i>	-	80	-
<i>Scopulariopsis/Microascus</i>	-	-	-
<i>Stachybotrys/Memnoniella</i>	-	-	-
<i>Unidentifiable Spores</i>	-	-	-
<i>Zygomycetes</i>	-	-	-
<i>Nigrospora</i>	-	10	-
<i>Hyphal Fragment</i>	-	-	-
<i>Insect Fragment</i>	-	80	-
<i>Pollen</i>	-	-	-
Total Fungi	1390	3410	No Trace

*Spore Counts per cubic meter of air (Counts/m³).

++Includes other spores with similar morphology.

Findings and Conclusions

The comfort parameters (i.e., temperature, RH, CO₂, and CO levels) in the representative areas conform to ASHRAE and/or NAAQS guidelines. On November 19, 2020, total mold counts in representative area samples (spore count/m³ of air) in all the areas inspected were lower than the outdoor concentrations, indicating no amplified mold growth.

Thank you for the opportunity to provide industrial hygiene services for PGCPs. If you have any questions, please contact me at 301.595.3783.

Sincerely,



Chaminda Jayatilake, PE, CIH, CSP, CHMM
Certified Industrial Hygienist
Soil and Land Use Technology Inc. (SaLUT)

Attachment

Attachment - Mold Spore Sample Analytical Results and Chain-of-Custody Forms

Attachment

Mold Spore Sample Analytical Results and Chain-of-Custody Forms



EMSL Analytical, Inc.

10768 Baltimore Avenue Beltsville, MD 20705

Tel/Fax: (301) 937-5700 / (301) 937-5701

<http://www.EMSL.com> / beltsvillelab@emsl.com

EMSL Order: 192011553

Customer ID: SALU50

Customer PO:

Project ID:

Attention: Indika Jayatilake

SaLUT

1818 New York Avenue, NE

Suite 231

Washington, DC 20002

Project: Green Valley Offices/ PGCPs IAQ

Phone: (301) 595-3783

Fax: (301) 595-3787

Collected Date: 11/19/2020

Received Date: 11/20/2020 08:30 AM

Analyzed Date: 11/24/2020

Test Report: Air-O-Cell™ Analysis of Fungal Spores & Particulates by Optical Microscopy (Methods MICRO-SOP-201, ASTM D7391)

Lab Sample Number:	192011553-0001			192011553-0002			192011553-0003		
Client Sample ID:	01			02			03		
Volume (L):	75			75			75		
Sample Location:	Cafeteria			2nd floor H/W next to C/R 202			2nd floor H/W next to CTE S/R 207		
Spore Types	Raw Count	Count/M ³	% of Total	Raw Count	Count/M ³	% of Total	Raw Count	Count/M ³	% of Total
Alternaria (Ulocladium)	-	-	-	-	-	-	-	-	-
Ascospores	-	-	-	-	-	-	-	-	-
Aspergillus/Penicillium	-	-	-	-	-	-	3	100	25.6
Basidiospores	3	100	50	6	300	76.9	3	100	25.6
Bipolaris++	-	-	-	-	-	-	-	-	-
Chaetomium	-	-	-	-	-	-	-	-	-
Cladosporium	3	100	50	1	40	10.3	3	100	25.6
Curvularia	-	-	-	-	-	-	-	-	-
Epicoccum	-	-	-	-	-	-	-	-	-
Fusarium	-	-	-	-	-	-	-	-	-
Ganoderma	-	-	-	-	-	-	1	40	10.3
Myxomycetes++	-	-	-	1*	10*	2.6	1	40	10.3
Pithomyces++	-	-	-	-	-	-	-	-	-
Rust	-	-	-	-	-	-	-	-	-
Scopulariopsis/Microascus	-	-	-	-	-	-	-	-	-
Stachybotrys/Memnoniella	-	-	-	-	-	-	-	-	-
Unidentifiable Spores	-	-	-	1	40	10.3	-	-	-
Zygomycetes	-	-	-	-	-	-	-	-	-
Nigrospora	-	-	-	-	-	-	-	-	-
Pestalotia/Pestalotiopsis	-	-	-	-	-	-	-	-	-
Torula-like	-	-	-	-	-	-	1*	10*	2.6
Total Fungi	6	200	100	9	390	100	12	390	100
Hyphal Fragment	-	-	-	2	80	-	-	-	-
Insect Fragment	-	-	-	-	-	-	1*	10*	-
Pollen	-	-	-	-	-	-	-	-	-
Analyt. Sensitivity 600x	-	42	-	-	42	-	-	42	-
Analyt. Sensitivity 300x	-	13*	-	-	13*	-	-	13*	-
Skin Fragments (1-4)	-	2	-	-	2	-	-	2	-
Fibrous Particulate (1-4)	-	1	-	-	1	-	-	1	-
Background (1-5)	-	1	-	-	1	-	-	1	-

++ Includes other spores with similar morphology; see EMSL's fungal glossary for each specific category.

Abubakar Barry, Microbiology Laboratory Manager
or other Approved Signatory

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Samples analyzed by EMSL Analytical, Inc. Plymouth Meeting, PA AIHA-LAP, LLC EMLAP #178659

Initial report from: 11/25/2020 08:19 AM

For information on the fungi listed in this report, please visit the Resources section at www.emsl.com



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Project: Green Valley Offices/ PGCPs IAQ

Phone: (301) 595-3783

Fax: (301) 595-3787

Collected Date: 11/19/2020

Received Date: 11/20/2020 08:30 AM

Analyzed Date: 11/24/2020

Test Report: Air-O-Cell™ Analysis of Fungal Spores & Particulates by Optical Microscopy (Methods MICRO-SOP-201, ASTM D7391)

Lab Sample Number: Client Sample ID: Volume (L): Sample Location:	192011553-0004			192011553-0005			192011553-0006		
	Raw Count	Count/M ³	% of Total	Raw Count	Count/M ³	% of Total	Raw Count	Count/M ³	% of Total
04 75 H/W next to Staff Lounge									
05 75 H/W next to C/R 404									
06 75 Outside Exterior EV Sample									
Spore Types									
Alternaria (Ulocladium)	-	-	-	-	-	-	-	-	-
Ascospores	-	-	-	-	-	-	3	100	2.9
Aspergillus/Penicillium	5	200	29.9	16	680	48.9	6	300	8.8
Basidiospores	10	420	62.7	12	510	36.7	25	1100	32.3
Bipolaris++	-	-	-	-	-	-	1	40	1.2
Chaetomium	-	-	-	-	-	-	-	-	-
Cladosporium	1	40	6	2	80	5.8	22	930	27.3
Curvularia	-	-	-	-	-	-	-	-	-
Epicoccum	-	-	-	-	-	-	2	80	2.3
Fusarium	-	-	-	-	-	-	-	-	-
Ganoderma	-	-	-	-	-	-	-	-	-
Myxomycetes++	1*	10*	1.5	2	80	5.8	16	680	19.9
Pithomyces++	-	-	-	1	40	2.9	-	-	-
Rust	-	-	-	-	-	-	2	80	2.3
Scopulariopsis/Microascus	-	-	-	-	-	-	-	-	-
Stachybotrys/Memnoniella	-	-	-	-	-	-	-	-	-
Unidentifiable Spores	-	-	-	-	-	-	-	-	-
Zygomycetes	-	-	-	-	-	-	-	-	-
Nigrospora	-	-	-	-	-	-	1*	10*	0.3
Pestalotia/Pestalotiopsis	-	-	-	-	-	-	1*	10*	0.3
Torula-like	-	-	-	-	-	-	2	80	2.3
Total Fungi	17	670	100	33	1390	100	81	3410	100
Hyphal Fragment	-	-	-	-	-	-	-	-	-
Insect Fragment	-	-	-	-	-	-	2	80	-
Pollen	-	-	-	-	-	-	-	-	-
Analyt. Sensitivity 600x	-	42	-	-	42	-	-	42	-
Analyt. Sensitivity 300x	-	13*	-	-	13*	-	-	13*	-
Skin Fragments (1-4)	-	2	-	-	2	-	-	1	-
Fibrous Particulate (1-4)	-	1	-	-	1	-	-	1	-
Background (1-5)	-	1	-	-	1	-	-	1	-

++ Includes other spores with similar morphology; see EMSL's fungal glossary for each specific category.

Abubakar Barry, Microbiology Laboratory Manager
or other Approved Signatory

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Samples analyzed by EMSL Analytical, Inc. Plymouth Meeting, PA AIHA-LAP, LLC EMLAP #178659

Initial report from: 11/25/2020 08:19 AM

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Attention: Indika Jayatilake

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1818 New York Avenue, NE

Suite 231

Washington, DC 20002

Project: Green Valley Offices/ PGCPs IAQ

Phone: (301) 595-3783

Fax: (301) 595-3787

Collected Date: 11/19/2020

Received Date: 11/20/2020 08:30 AM

Analyzed Date: 11/24/2020

Test Report: Air-O-Cell™ Analysis of Fungal Spores & Particulates by Optical Microscopy (Methods MICRO-SOP-201, ASTM D7391)

Lab Sample Number:	192011553-0007		
Client Sample ID:	07		
Volume (L):			
Sample Location:	Field Blank		
Spore Types	Raw Count	Count/M³	% of Total
Alternaria (Ulocladium)	-	-	-
Ascospores	-	-	-
Aspergillus/Penicillium	-	-	-
Basidiospores	-	-	-
Bipolaris++	-	-	-
Chaetomium	-	-	-
Cladosporium	-	-	-
Curvularia	-	-	-
Epicoccum	-	-	-
Fusarium	-	-	-
Ganoderma	-	-	-
Myxomycetes++	-	-	-
Pithomyces++	-	-	-
Rust	-	-	-
Scopulariopsis/Microascus	-	-	-
Stachybotrys/Memnoniella	-	-	-
Unidentifiable Spores	-	-	-
Zygomycetes	-	-	-
Nigrospora	-	-	-
Pestalotia/Pestalotiopsis	-	-	-
Torula-like	-	-	-
Total Fungi	-	No Trace	-
Hyphal Fragment	-	-	-
Insect Fragment	-	-	-
Pollen	-	-	-
Analyt. Sensitivity 600x	-	0	-
Analyt. Sensitivity 300x	-	0*	-
Skin Fragments (1-4)	-	-	-
Fibrous Particulate (1-4)	-	-	-
Background (1-5)	-	-	-

++ Includes other spores with similar morphology; see EMSL's fungal glossary for each specific category.

Abubakar Barry, Microbiology Laboratory Manager
or other Approved Signatory

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EMSL ANALYTICAL, INC.
LABORATORY PRODUCTS TRAINING

Microbiology Chain of Custody

EMSL Order Number (Lab Use Only):

192011553

PHONE:

FAX:

Company Name: SaLUT Inc.		EMSL-Bill to: <input checked="" type="checkbox"/> Same <input type="checkbox"/> Different If Bill to is Different note instructions in Comments**					
Street: 1818 New York Ave NE Suite 231		Third Party Billing requires written authorization from third party					
City: Washington	State/Province: DC	Zip/Postal Code: 20002	Country: USA				
Report To (Name): Indika Jayatilake		Telephone #: 301-595-3783					
Email Address: ijayatilake@salutinc.com		Fax #:	Purchase Order:				
Project Number/Location: Green Valley Offices / PGCPs IAQ		Please Provide Results: <input type="checkbox"/> Fax <input checked="" type="checkbox"/> Email					
Location Address: 2215 Chadwick St, Hillcrest Heights, MD 20748		Connecticut Samples: <input type="checkbox"/> Commercial <input type="checkbox"/> Residential					
*Analysis completed in accordance with EMSL's Terms and Conditions located in the Analytical Price Guide. TATs are subject to methodology requirements							
Sterile, Sodium Thiosulfate Preserved Bottle Used: <input type="checkbox"/> Biocide Used in Source (specify): <input type="checkbox"/>							
Public Water Supply Samples: <input type="checkbox"/> Note: All results may automatically be reported to DOH if required by state.							
Turnaround Time (TAT) Options * - Please Check							
<input type="checkbox"/> 3 Hour	<input type="checkbox"/> 6 Hour	<input type="checkbox"/> 24 Hour	<input type="checkbox"/> 48 Hour	<input checked="" type="checkbox"/> 72 Hour	<input type="checkbox"/> 96 Hour	<input type="checkbox"/> 1 Week <input type="checkbox"/> 2 Week	
Microbiology Test Codes							
M001 Air-O-Cell.	M174 MoldSnap	M024 Pseudomonas aeruginosa (MFT*)	M115 Sewage Screen - Water (P/A***)				
M030 Micro 5	M032 Allergenco-D	M015 Heterotrophic Plate Count	M116 Sewage Screen - Water (MPN**)				
M041 Fungal Direct Examination		M017 Total Coliform & E. coli (Colilert P/A***)	M117 Sewage Screen - Swab (P/A***)				
M169 Pollen ID & Enumeration		M018 Total Coliform & E. coli (MFT*)	M013 Sewage Screen - Swab (MFT*)				
M280 Dust Characterization Level-1		M114 Total Coliform & E. coli Enumeration (Colilert MPN**)	M133 Methicillin-resistant Staph. aureus (MRSA)				
M281 Dust Characterization Level-2		M019 Fecal Coliform (MFT*)	M031 Rapid-growing non-TB Mycobacteria Detection & Enumeration				
M005 Viable Fungi- Air Samples (Genus ID & Count)		M020 Fecal Streptococcus (MFT*)	M014 Endotoxin Analysis				
M006 Viable Fungi- Air Samples (Includes Penicillium, Aspergillus, Cladosporium, Stachybotrys Species ID & Count)		M029 Enterococci (MFT*)	M044 Group Allergen (Cat, Dog, Cockroach, Dust Mite)				
M007 Culturable fungi - Surface Samples (Genus ID & Count)		M129 Enterococci (Enterolert P/A***)	Other See Analytical Price Guide				
M008 Culturable fungi - Surface Samples (Includes Penicillium, Aspergillus, Cladosporium, Stachybotrys Species ID & Count)		M180 Real Time qPCR-ERMI 36 Panel	Legionella Analysis Please use EMSL Legionella COC				
M009 Bacteria Culture Gram Stain & Count		M025 Sewage Screen -Water (MFT*)					
M010 Bacteria Count & ID - 3 Most Prominent		*MFT= Membrane Filtration Technique					
M011 Bacteria Count & ID - 5 Most Prominent		**MPN= Most Probable Number					
M012 Pseudomonas aeruginosa (P/A***)		***P/A= Presence/Absence					
Name of Sampler: Jude Fonseka		Signature of Sampler:					
Sample #	Sample Location/Description	Sample Type	Potable/ NonPotable (only for waters)	Test Code	Volume/ Area	Date/Time Collected	Temperature (°C) (Lab Use Only)
01	Cafeteria	Air		M001	75L	11/19/2020	
02	2nd floor H/W next to C/R 202	Air		M001	75L	11/19/2020	
03	2nd floor H/W next to CTE S/R 207	Air		M001	75L	11/19/2020	
04	H/W next to Staff Lounge	Air		M001	75L	11/19/2020	
05	H/W next to C/R 404	Air		M001	75L	11/19/2020	
06	Outside Exterior EV Sample	Air		M001	75L	11/19/2020	
Client Sample # (s): -		Total # of Samples: 07		Samples Received Chilled? Yes / No			
Relinquished (Client):		Date:		Time:		RECEIVED EMSL ANALYTICAL, INC. BELTSVILLE, MD 2020 NOV 20 A 6:14	
Received (Lab):		Date:		Time:			
Comments/Special Instructions:							

