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June 9, 2019

Prince George's County Public Schools  
13300 Old Marlboro Pike  
Upper Marlboro, Maryland 20772  
Attention: Mr. Alex Baylor

RE: Indoor Air Quality Screening, James McHenry Elementary School  
IFB: 022-19  
ATI Project Number: ATI19-663

Dear Mr. Baylor:

Prince George's County Public Schools requested that ATI, Inc., conduct a proactive indoor air quality (IAQ) screening at James McHenry Elementary School. The IAQ screening was conducted on May 15, 2019. Its key findings are enclosed in the Executive Summary on page three, and the official laboratory report for total fungal spore trap sampling is enclosed in Appendix A.

Thank you for the opportunity to provide Industrial Hygiene services for Prince George's County Public Schools. If you have any questions regarding this report, please contact us at (202) 643-4283.

Sincerely,  
**ATI, INC.**

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Courtney E. McCall  
Project Manager

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Sarath Seneviratne  
CIH, CSP, CHMM

# Indoor Air Quality Screening Report



Prince George's County Public Schools  
James McHenry Elementary School  
8909 McHenry Lane  
Lanham, Maryland 20706

Prepared for:

Prince George's County Public Schools  
13300 Old Marlboro Pike  
Upper Marlboro, Maryland 20772

**June 9, 2018**

Submitted by:

The logo for ATI (Air Technology, Inc.) consists of the lowercase letters "ati" in a bold, blue, sans-serif font. The letters are positioned centrally on the page.

ATI Job # 19-663

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Table of Contents

1. Executive Summary and Key Findings ..... 3

2. Assessment Methods ..... 3

3. Visual Observations..... 4

4. Thermal Environmental Conditions for Human Occupancy ..... 5

    4.1 Temperature..... 5

    4.2 Relative Humidity ..... 5

    4.3 Carbon Dioxide ..... 6

    4.4 Carbon Monoxide..... 7

5. Total Fungal Air Sampling Results ..... 7

6. Summary of Findings ..... 8

Table 1: Visual Observations and Sampling Locations..... 4

Table 2: Temperature Measurements ..... 5

Table 3: Relative Humidity Measurements..... 6

Table 4: Carbon Dioxide Measurements..... 6

Table 5: Carbon Monoxide Measurements ..... 7

Appendix A: Laboratory Report and Chain of Custody  
Appendix B: Instrument Calibration Records

### Abbreviations and Acronyms

<b>AHU</b>	Air-Handling Unit
<b>AIHA</b>	American Industrial Hygiene Association
<b>ASHRAE</b>	American Society of Heating, Refrigerating and Air-Conditioning Engineers
<b>ASTM</b>	American Society for Testing and Materials
<b>CO</b>	Carbon Monoxide
<b>CO<sub>2</sub></b>	Carbon Dioxide
<b>EMLAP</b>	Environmental Microbiology Laboratory Accreditation Program
<b>HVAC</b>	Heating, Ventilating, And Air-Conditioning
<b>IAQ</b>	Indoor Air Quality
<b>NIST</b>	National Institute for Standards and Technology
<b>NVLAP</b>	National Voluntary Laboratory Accreditation Program
<b>RH</b>	Relative Humidity

Abbreviations involving scientific volume and measurements involving media or water sampling

<b>Counts/m<sup>3</sup></b>	Mold spores per cubic meter of air
<b>LPM</b>	Liters Per Minute
<b>NTE</b>	Not to exceed

## 1. Executive Summary and Key Findings

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ATI conducted a proactive Indoor Air Quality (IAQ) screening on May 15, 2019, at James McHenry Elementary School, located at 8909 McHenry Lane, Lanham, MD 20706.

The screening included a visual assessment of randomly selected classrooms and other frequently occupied spaces, such as the cafeteria, the main office, and classrooms, for potential IAQ contributors and pathways. As part of the screening, ATI collected direct reading measurements for comfort parameters, including temperature, relative humidity, carbon dioxide, and carbon monoxide. Also, ATI collected total fungal air samples on spore trap cassettes for microbiological analysis.

The following is a summary of the key findings from this screening:

1. Temperature measurements were slightly below and on the lower end of ASHRAE guidelines for summer temperatures, between 73°F and 79°F;
2. Humidity measurements were within ASHRAE guidelines, <65%.
3. Carbon dioxide measurements in two locations exceeded the ASHRAE maximum for the day of the screening, 1,100 parts per million (ppm).
4. Carbon monoxide was not detected throughout the tested spaces.
5. Laboratory analysis indicated that total fungal concentrations on the spore traps did not show significant indoor amplification.

## 2. Assessment Methods

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Mr. Brian Chapman and Ms. Mikal Frater of ATI, Inc., conducted a visual assessment and air sampling on May 15, 2019. Sampled rooms were randomly selected and accounted for approximately 10% of classrooms. Visual observations were made at the time the samples were collected. ATI references the American Society of Heating, Refrigerating, and Air-Conditioning Engineers (ASHRAE) *Standard 62.1 – 2016* and *ASHRAE Standard 55 – 2017* when providing IAQ services to clients. ASHRAE is an industry leader on energy efficiency and indoor air quality.

All measurements and air samples were collected between three-six feet from floor elevation, which represents the breathing zone, and away from air-supply and return diffusers. Real-time direct readings for temperature, relative humidity, carbon dioxide (CO<sub>2</sub>), and carbon monoxide (CO), were obtained with a calibrated TSI Q-Trak 7575-X Meter and attached 982 Probe.

Total fungal air samples were collected with a Buck BioAire High-Volume Sampling Pump on Zefon Air-O-Cell spore-trap cassettes at a flow rate of 15 liters per minute for five minutes, for a sample volume of 75 liters. The samples were analyzed by direct microscopic examination (identifies and counts both viable and non-viable spores, which is then considered “total fungal”), via the American Society for Testing and Materials (ASTM) Standard D7391-09 by EMSL Analytical, Inc., (EMSL) located in Beltsville, MD.

EMSL participates in the National Institute of Standards and Technology’s (NIST’s) National Voluntary Laboratory Accreditation Program (NVLAP) for general laboratory performance and management and the

American Industrial Hygiene Association (AIHA) Environmental Microbial Laboratory Accreditation Program (EMLAP, Certificate Number 102891).

Instrument calibration records are included in Appendix B of this report.

**3. Visual Observations**

**Table 1: Visual Observations and Sampling Locations**

Sample Location	Observations
Outside	<ul style="list-style-type: none"> <li>• Clear skies.</li> <li>• Parking lot surrounded by trees.</li> <li>• No vehicle or foot traffic.</li> <li>• Sunny with light winds.</li> <li>• Eight air handling units can be seen on rooftop from outside.</li> </ul>
Main Office	<ul style="list-style-type: none"> <li>• Two wall units.</li> <li>• One air diffuser with light dirt accumulation.</li> <li>• Five occupants in room during sampling.</li> <li>• Light foot traffic.</li> <li>• One printer/fax machine about 10 ft. from sampling area.</li> <li>• Individual fan in room – OFF.</li> <li>• Space is approximately 390 ft.<sup>2</sup></li> </ul>
Room 8	<ul style="list-style-type: none"> <li>• One long wall unit along wall entirely.</li> <li>• Three occupants in room during sampling.</li> <li>• Very light foot traffic.</li> <li>• Friedrich A/C unit in room.</li> <li>• Heat is on in boiler system.</li> <li>• Space is approximately 750 ft.<sup>2</sup></li> </ul>
Room 1	<ul style="list-style-type: none"> <li>• Labeled “computer lab” on floor plan.</li> <li>• Friedrich A/C unit (SL36L30A-C).</li> <li>• One long wall unit along south perimeter wall.</li> <li>• Emergency exit in room – outside access.</li> <li>• A/C unit is installed but not properly insulated – breach allowing outside air into space.</li> <li>• Filter is clean, but coils have trace dirt accumulation.</li> <li>• Space is approximately 720 ft.<sup>2</sup></li> </ul>
Room A4	<ul style="list-style-type: none"> <li>• Twenty-one occupants in area during sampling, about six feet from sampling area.</li> <li>• Water stain on ceiling tile indicative of previous leak in room.</li> <li>• Four air diffusers, one air return.</li> <li>• Air return 12x12 pleated filter with mild dirt load.</li> <li>• Space is approximately 820 ft.<sup>2</sup></li> </ul>
Room 19	<ul style="list-style-type: none"> <li>• Wall unit along wall perimeter.</li> <li>• Two window A/C units.</li> <li>• About seventeen occupants in room during sampling.</li> <li>• Bathroom within classroom.</li> <li>• Hot water non-operable (per teacher).</li> </ul>

Sample Location	Observations
	<ul style="list-style-type: none"> <li>Water leaks from toilet base.</li> <li>Dripping faucet.</li> <li>Space is approximately 690 ft.<sup>2</sup></li> </ul>

#### 4. Thermal Environmental Conditions for Human Occupancy

ASHRAE Standard 55-2017, *Thermal Environmental Conditions for Human Occupancy*, addresses thermal comfort in an office environment, which means that an employee wearing a normal amount of clothing feels neither too cold nor too warm. This standard discusses thermal comfort within the context of air temperature, humidity, and air movement and provides recommended ranges for temperature and humidity that are intended to satisfy most building occupants. The recommended ASHRAE ranges are referenced below by each comfort parameter.

##### 4.1 Temperature

The ASHRAE standard establishes a winter comfort range of between 68°F and 75°F and a summer range of between 73°F and 79°F. The temperature measurements obtained during the May 15, 2019, screening is summarized in Table 2. As indicated by the data in the table, temperatures in the school averaged between 68.2 - 73.9°F, below and on the lower end of the ASHRAE summer comfort range.

Table 2: Temperature Measurements

Sample Location	May 15, 2019 °F			ASHRAE Standard °F
	Min	Max	Average	
Outside	58.0	58.0	58.0	N/A
<b>Indoors</b>				
Main Office	73.8	74.0	73.9	73 – 79
Room 8	70.1	70.1	70.1	73 – 79
Room 1	69.3	69.3	69.3	73 – 79
Room A4	71.7	71.9	71.8	73 – 79
Room 19	67.8	68.5	68.2	73 – 79

##### 4.2 Relative Humidity

Relative humidity is a key factor for mold growth. Mold has the potential of growing on suitable surfaces with humidity levels above 65%. ASHRAE Standard 62.1-2016, *Ventilation for Acceptable Indoor Air Quality*, recommends a maximum indoor relative humidity of 65% to prevent condensation of moisture on surfaces. Relative humidity below 30% may result in drying of the mucous membranes and skin. Relative humidity measurements are summarized in Table 3. As indicated by the data in the table, relative humidity measurements averaged between 41.5 and 49.4%, below the ASHRAE maximum recommendation of 65% relative humidity.



Table 3: Relative Humidity Measurements

Sample Location	May 15, 2019 (%)			ASHRAE Standard (% RH)
	Min	Max	Average	
Outside	33.0	35.1	34.05	N/A
<b>Inside</b>				
Main Office	41.3	41.7	41.5	< 65
Room 8	48.1	48.2	48.2	< 65
Room 1	46.0	46.8	46.4	< 65
Room A4	48.4	48.8	48.6	< 65
Room 19	48.8	49.9	49.4	< 65

### 4.3 Carbon Dioxide

Carbon dioxide measurements within an occupied building are a standard method used to gauge the efficiency of ventilation systems. Carbon dioxide is a by-product of human respiration and does not pose an acute health hazard alone. Elevated concentrations may suggest that insufficient fresh air is being supplied to an occupied space and/or that the ventilation system does not provide a sufficient rate of air exchange.

Research has indicated that buildings with adequately operating ventilation systems are able to remove odors generated by activities in an indoor office environment efficiently. ASHRAE *Standard 62.1-2016* states that comfort (odor) criteria with respect to human bioeffluents are likely to be satisfied if the ventilation results indoor carbon dioxide concentrations are less than 700 parts per million (ppm) above the outdoor air concentration.

Carbon dioxide measurements are summarized in Table 4. On the day of the screening, the average outdoor carbon dioxide concentration obtained was 400 ppm, which calculates to a maximum indoor concentration of 1,100 ppm (700 + 400). The carbon dioxide levels inside the school ranged from the average minimum detected, 935.5 ppm to 1,366 ppm, the average maximum detected, with two locations exceeding the maximum recommended concentration of 1,100 ppm.

Table 4: Carbon Dioxide Measurements

Sample Location	May 15, 2019 Concentration (parts per million)			ASHRAE Standard (ppm) NTE
	Min	Max	Average	
Outside	396	404	400	N/A
<b>Inside</b>				
Main Office	965	972	968.5	1,100
Room 8	1,365	1,367	1,366	1,100
Room 1	955	961	958	1,100
Room A4	1,204	1,228	1,216	1,100
Room 19	925	946	935.5	1,100

#### 4.4 Carbon Monoxide

Carbon monoxide is a colorless and odorless gas produced by the incomplete combustion of carbon containing fuels. Oil, gasoline, diesel fuels, wood, coke, and coal are the major sources of carbon monoxide. ASHRAE recommends that carbon monoxide not exceed nine ppm indoors. As indicated by the data in Table 5, carbon monoxide was not detected throughout the school.

**Table 5: Carbon Monoxide Measurements**

Sample Location	May 15, 2019 Concentration (parts per million)			ASHRAE Standard (ppm)
	Min	Max	Average	
Outside	0	0	0	N/A
<b>Inside</b>				
Main Office	0	0	0	< 9
Room 8	0	0	0	< 9
Room 1	0	0	0	< 9
Room A4	0	0	0	< 9
Room 19	0	0	0	< 9

## 5. Total Fungal Air Sampling Results

Mold needs a food source, moisture, proper temperature and humidity, and at most times, a source of light, to grow in an environment. Air filtration through building entrances and exits, open windows and loading docks, and foot traffic into buildings, serve as primary pathways that bring mold indoors. Water leaks and humid conditions inside of buildings provide the moisture that fosters mold growth.

The May 15, 2019, mold screening sampled air using spore trap cassettes in randomly selected classrooms and other rooms. These cassettes collect both viable spores, those capable of producing more fungal colonies, and non-viable spores, which cannot reproduce. Based upon recognized industry practices, indoor mold concentrations are compared with those detected outdoors, which are also known as ambient or baseline samples.

In normal circumstances, the diversity of spores identified indoors and outdoors should be similar with some exceptions. The dominating presence of one or two species of fungal spores identified indoors and the absence of the same species outdoors can indicate a moisture problem with the potential to degrade the air quality. Fungi species present indoors are typically found at levels ranging from approximately 10-50% of their levels in the outdoor air, reflecting the filtering by the building's HVAC system.

The official laboratory report with spore trap samples collected on May 15, 2019, is presented in Appendix A. The findings indicated that the indoor concentrations were favorable compared to the outdoor concentrations, and indoor amplification was not present. Total concentrations detected in each tested space did not exceed the spore counts detected outdoors, 13,080 counts/m<sup>3</sup>.

The laboratory report indicates that Basidiospores are the highest concentration within the samples. Basidiospores are common outdoor fungi with the potential to enter building spaces through main entrances, opened windows, or via building envelopes. They may pose allergy-like symptoms but are not fungi to be associated with water damaged building materials. Basidiospores detected indoors did not exceed the quantity detected outdoors. Also, Ascospores and Cladosporium, spores commonly found indoors and known to cause allergies, were detected in most tested rooms. Aspergillus/Penicillium, which are known to cause allergies also, were detected in four of the six tested spaces.

## 6. Summary of Findings

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ASHRAE comfort parameters including relative humidity and carbon monoxide were within recommended ranges in all tested areas. The indoor temperatures fell below and on the lower end of the ASHRAE recommended summer comfort range. Carbon dioxide levels in two locations exceeded the ASHRAE maximum for the day of the screening.

Generally, total indoor concentrations of mold compared favorably to those detected outdoors, with no locations exceeding the total quantity detected outdoors.

We appreciate the opportunity to provide these IAQ testing services for you. If you have any questions, please contact us at (202) 643-4283.

Best regards,  
**ATI, INC.**



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Courtney E. McCall  
Project Manager



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Sarath Seneviratne  
CIH, CSP, CHMM

**Appendix A:  
Laboratory Report and Chain of Custody**



# EMSL Analytical, Inc.

2500 Gateway Centre Blvd., Suite 600 Morrisville, NC 27560

Tel/Fax: (919) 465-3900 / (919) 465-3950

<http://www.EMSL.com> / [rleighlab@emsl.com](mailto:rleighlab@emsl.com)

<b>EMSL Order:</b> 291905208
<b>Customer ID:</b> ATII25A
<b>Customer PO:</b>
<b>Project ID:</b>

<b>Attn:</b> Brian Chapman ATI 4221 Forbes Blvd Suite 250 Lanham, MD 20706 <b>Project:</b> 19-663 - PGCPs - McHenry ES	<b>Phone:</b> (202) 368-1376 <b>Fax:</b> <b>Collected:</b> 05/15/2019 <b>Received:</b> 05/15/2019 <b>Analyzed:</b> 05/20/2019
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### Test Report: Air-O-Cell™ Analysis of Fungal Spores & Particulates by Optical Microscopy (Methods MICRO-SOP-201, ASTM D7391)

Lab Sample Number: Client Sample ID: Volume (L): Sample Location	291905208-0001 19-663-01 75 Outside Parking Lot			291905208-0002 19-663-02 Field Blank			291905208-0003 19-663-03 75 Main Office			
	Spore Types	Raw Count	Count/m³	% of Total	Raw Count	Count/m³	% of Total	Raw Count	Count/m³	% of Total
Alternaria (Ulocladium)	-	-	-	-	-	-	-	-	-	-
Ascospores	67	2800	21.4	-	-	-	7	300	21.4	
Aspergillus/Penicillium	-	-	-	-	-	-	15	630	45	
Basidiospores	241	10200	78	-	-	-	7	300	21.4	
Bipolaris++	-	-	-	-	-	-	-	-	-	
Chaetomium	-	-	-	-	-	-	-	-	-	
Cladosporium	2	80	0.6	-	-	-	3	100	7.1	
Curvularia	-	-	-	-	-	-	-	-	-	
Epicoccum	-	-	-	-	-	-	-	-	-	
Fusarium	-	-	-	-	-	-	-	-	-	
Ganoderma	-	-	-	-	-	-	-	-	-	
Myxomycetes++	-	-	-	-	-	-	4*	50*	3.6	
Pithomyces++	-	-	-	-	-	-	1*	10*	0.7	
Rust	-	-	-	-	-	-	1*	10*	0.7	
Scopulariopsis/Microascus	-	-	-	-	-	-	-	-	-	
Stachybotrys/Memnoniella	-	-	-	-	-	-	-	-	-	
Unidentifiable Spores	-	-	-	-	-	-	-	-	-	
Zygomycetes	-	-	-	-	-	-	-	-	-	
<b>Total Fungi</b>	<b>310</b>	<b>13080</b>	<b>100</b>	-	<b>No Trace</b>	-	<b>38</b>	<b>1400</b>	<b>100</b>	
Hyphal Fragment	-	-	-	-	-	-	1	40	-	
Insect Fragment	-	-	-	-	-	-	-	-	-	
Pollen	1	40	-	-	-	-	3	100	-	
Analyt. Sensitivity 600x	-	42	-	-	0	-	-	42	-	
Analyt. Sensitivity 300x	-	13*	-	-	0*	-	-	13*	-	
Skin Fragments (1-4)	-	1	-	-	-	-	-	3	-	
Fibrous Particulate (1-4)	-	-	-	-	-	-	-	1	-	
Background (1-5)	-	1	-	-	-	-	-	4	-	

++ Includes other spores with similar morphology; see EMSL's fungal glossary for each specific category.

*Alan Goldstein*

Alan Goldstein, Ph.D., Laboratory Manager  
or other approved signatory

High levels of background particulate can obscure spores and other particulates leading to underestimation. Background levels of 5 indicate an overloading of background particulates, prohibiting accurate detection and quantification. Present = Spores detected on overloaded samples. Results are not blank corrected unless otherwise noted. The detection limit is equal to one fungal spore, structure, pollen, fiber particle or insect fragment. \*\*\* Denotes particles found at 300X. "-" Denotes not detected. Due to method stopping rules, raw counts in excess of 100 are extrapolated based on the percentage analyzed. EMSL maintains liability limited to cost of analysis. This report relates only to the samples reported above and may not be reproduced, except in full, without written approval by EMSL. EMSL bears no responsibility for sample collection activities or analytical method limitations. Interpretation and use of test results are the responsibility of the client. Samples received in good condition unless otherwise noted.

Samples analyzed by EMSL Analytical, Inc. Morrisville, NC AIHA-LAP, LLC--EMLAP Lab 173741

Initial report from: 05/21/2019 08:12:10

For information on the fungi listed in this report, please visit the Resources section at [www.emsl.com](http://www.emsl.com)



# EMSL Analytical, Inc.

2500 Gateway Centre Blvd., Suite 600 Morrisville, NC 27560

Tel/Fax: (919) 465-3900 / (919) 465-3950

<http://www.EMSL.com> / [raleighlab@emsl.com](mailto:raleighlab@emsl.com)

EMSL Order: 291905208

Customer ID: ATII25A

Customer PO:

Project ID:

Attn: Brian Chapman

ATI

4221 Forbes Blvd

Suite 250

Lanham, MD 20706

Project: 19-663 - PGCPs - McHenry ES

Phone: (202) 368-1376

Fax:

Collected: 05/15/2019

Received: 05/15/2019

Analyzed: 05/20/2019

### Test Report: Air-O-Cell™ Analysis of Fungal Spores & Particulates by Optical Microscopy (Methods MICRO-SOP-201, ASTM D7391)

Lab Sample Number:	291905208-0004			291905208-0005			291905208-0006		
Client Sample ID:	19-663-04			19-663-05			19-663-06		
Volume (L):	75			75			75		
Sample Location	Room 8			Computer Lab Room 1			Room A4		
Spore Types	Raw Count	Count/m <sup>3</sup>	% of Total	Raw Count	Count/m <sup>3</sup>	% of Total	Raw Count	Count/m <sup>3</sup>	% of Total
Alternaria (Ulocladium)	-	-	-	-	-	-	-	-	-
Ascospores	12	510	28	56	2400	52.7	5	200	31.3
Aspergillus/Penicillium	7	300	16.5	1	40	0.9	-	-	-
Basidiospores	11	460	25.3	37	1600	35.2	4	200	31.3
Bipolaris++	-	-	-	-	-	-	-	-	-
Chaetomium	-	-	-	-	-	-	-	-	-
Cladosporium	12	510	28	12	510	11.2	4	200	31.3
Curvularia	-	-	-	-	-	-	-	-	-
Epicoccum	-	-	-	-	-	-	1	40	6.3
Fusarium	-	-	-	-	-	-	-	-	-
Ganoderma	-	-	-	-	-	-	-	-	-
Myxomycetes++	1	40	2.2	-	-	-	-	-	-
Pithomyces++	-	-	-	-	-	-	-	-	-
Rust	-	-	-	-	-	-	-	-	-
Scopulariopsis/Microascus	-	-	-	-	-	-	-	-	-
Stachybotrys/Memnoniella	-	-	-	-	-	-	-	-	-
Unidentifiable Spores	-	-	-	-	-	-	-	-	-
Zygomycetes	-	-	-	-	-	-	-	-	-
<b>Total Fungi</b>	<b>43</b>	<b>1820</b>	<b>100</b>	<b>106</b>	<b>4550</b>	<b>100</b>	<b>14</b>	<b>640</b>	<b>100</b>
Hyphal Fragment	9	400	-	1*	10*	-	-	-	-
Insect Fragment	-	-	-	-	-	-	-	-	-
Pollen	1*	10*	-	-	-	-	2*	30*	-
Analyt. Sensitivity 600x	-	42	-	-	42	-	-	42	-
Analyt. Sensitivity 300x	-	13*	-	-	13*	-	-	13*	-
Skin Fragments (1-4)	-	2	-	-	3	-	-	3	-
Fibrous Particulate (1-4)	-	1	-	-	1	-	-	1	-
Background (1-5)	-	2	-	-	1	-	-	2	-

++ Includes other spores with similar morphology; see EMSL's fungal glossary for each specific category.

Alan Goldstein, Ph.D., Laboratory Manager  
or other approved signatory

High levels of background particulate can obscure spores and other particulates leading to underestimation. Background levels of 5 indicate an overloading of background particulates, prohibiting accurate detection and quantification. Present = Spores detected on overloaded samples. Results are not blank corrected unless otherwise noted. The detection limit is equal to one fungal spore, structure, pollen, fiber particle or insect fragment. \*\*\* Denotes particles found at 300X. "-" Denotes not detected. Due to method stopping rules, raw counts in excess of 100 are extrapolated based on the percentage analyzed. EMSL maintains liability limited to cost of analysis. This report relates only to the samples reported above and may not be reproduced, except in full, without written approval by EMSL. EMSL bears no responsibility for sample collection activities or analytical method limitations. Interpretation and use of test results are the responsibility of the client. Samples received in good condition unless otherwise noted.

Samples analyzed by EMSL Analytical, Inc. Morrisville, NC AIHA-LAP, LLC--EMLAP Lab 173741

Initial report from: 05/21/2019 08:12:10

For information on the fungi listed in this report, please visit the Resources section at [www.emsl.com](http://www.emsl.com)

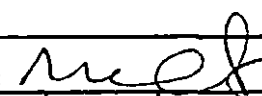


Client: ATI  
 Order: 291905208  
 Disposition: Discard after 7/14/2019

Test: M001 Air-O-Cell #Samples: 7  
 Project: 19-663 - PGCPs - McHenry ES

ATL, INC.  
 NORTH  
 108077  
 J-3675  
 -0262

EMSL Analytical, Inc.  
 LABORATORY PRODUCTS TRAINING

Company Name: <b>ATI, Inc</b>			EMSL-Bill to: <input type="checkbox"/> Same <input type="checkbox"/> Different If Bill to is Different note instructions in Comments				
Street: 4221 Rumsey Road, Suite 250			Third Party Billing requires written authorization from third party.				
City: Lanham	State/Province: MD	Zip/Postal Code: 20706	Country:				
Report To (Name): Brian Chapman / Mikal Frater			Telephone #: 202-558-7489				
Email Address: Brian@atiin.com & Mikal@atiinc.com			Fax #:		Purchase Order:		
Project Name/Number: 19-663- PGCPs - McHenry ES			Please Provide Results: <input type="checkbox"/> Fax <input checked="" type="checkbox"/> Email				
U.S. State Samples Taken:		Project Zip Code:		Connecticut Samples: <input checked="" type="checkbox"/> Commercial <input type="checkbox"/> Residential			
Sterile, Sodium Thiosulfate Preserved Bottle Used: <input type="checkbox"/> Biocide Used in Source (specify): <input type="checkbox"/>							
Public Water Supply Samples: <input type="checkbox"/> Note: All results may automatically be reported to DOH if required by state.							
Turnaround Time (TAT) Options - Please Check							
<input type="checkbox"/> 3 Hour	<input type="checkbox"/> 6 Hour	<input type="checkbox"/> 24 Hour	<input type="checkbox"/> 48 Hour	<input type="checkbox"/> 72 Hour	<input type="checkbox"/> 96 Hour	<input checked="" type="checkbox"/> 1 Week <input type="checkbox"/> 2 Week	
<b>Microbiology Test Codes</b>							
M001 Air-O-Cell	M174 MoldSnap	M012 Pseudomonas aeruginosa (P/A***)	M115 Sewage Screen - Water (P/A***)				
M030 Micro 5	M032 Allergenco-D	M024 Pseudomonas aeruginosa (MFT*)	M116 Sewage Screen - Water (MPN**)				
M041 Fungal Direct Examination		M015 Heterotrophic Plate Count	M117 Sewage Screen - Swab (P/A***)				
M169 Pollen ID & Enumeration		M017 Total Coliform & E. coli (Colilert P/A***)	M013 Sewage Screen - Swab (MFT*)				
M280 Dust Characterization Level-1		M018 Total Coliform & E. coli (MFT*)	M133 Methicillin-resistant Staph. aureus (MRSA)				
M281 Dust Characterization Level-2		M114 Total Coliform & E. coli Enumeration (Colilert MPN**)	M031 Rapid-growing non-TB Mycobacteria Detection & Enumeration				
M005 Viable Fungi- Air Samples (Genus ID & Count)		M019 Fecal Coliform (MFT*)	M014 Endotoxin Analysis				
M006 Viable Fungi- Air Samples (Includes Penicillium, Aspergillus, Cladosporium, Stachybotrys Species ID & Count)		M020 Fecal Streptococcus (MFT*)	M044 Group Allergen (Cat, Dog, Cockroach, Dust Mite)				
M007 Culturable fungi - Surface Samples (Genus ID & Count)		M029 Enterococci (MFT*)	Other See Analytical Price Guide				
M008 Culturable fungi - Surface Samples (Includes Penicillium, Aspergillus, Cladosporium, Stachybotrys Species ID & Count)		M129 Enterococci (Enterolert P/A***)	Legionella Analysis Please use EMSL Legionella COC				
M009 Bacteria Culture Gram Stain & Count		M180 Real Time qPCR-ERMI 36 Panel					
M010 Bacteria Count & ID - 3 Most Prominent		M025 Sewage Screen -Water (MFT*)					
M011 Bacteria Count & ID - 5 Most Prominent							
			*MFT= Membrane Filtration Technique **MPN= Most Probable Number ***P/A= Presence/Absence				
Name of Sampler: Brian Chapman & Mikal Frater			Signature of Sampler: 				
Sample #	Sample Location/Description	Sample Type	Potable/ NonPotable (Only for Waters)	Test Code	Volume/ Area	Date/Time Collected	Temperature (°C) (Lab Use Only)
Example A1	Kitchen Sink/Tap	Water	<input checked="" type="checkbox"/> P <input type="checkbox"/> NP	M017	100 mL	9/1/13 4:00 PM	
19-663-01	Outside Parking Lot	Air	<input type="checkbox"/> P <input type="checkbox"/> NP	M001	75L	05-15-19 - 9:13	
19-663-02	Field Blank	Air	<input type="checkbox"/> P <input type="checkbox"/> NP	M001	75L	05-15-19 -	
19-663-03	Main Office	Air	<input type="checkbox"/> P <input type="checkbox"/> NP	M001	75L	05-15-19 - 9:28	
19-663-04	Room 8	Air	<input type="checkbox"/> P <input type="checkbox"/> NP	M001	75L	05-15-19 - 9:38	
19-663-05	"Computer Lab" Room 1	Air	<input type="checkbox"/> P <input type="checkbox"/> NP	M001	75L	05-15-19 - 9:48	
Client Sample # (s): - 7		Total # of Samples: 7		Samples Received Chilled? Yes / No (Lab Use Only)			
Relinquished (Client): <b>MIKAL FRATER</b>			Date: 5-15-19		Time: <b>3:55</b>		
Received (Lab): <i>L. Bernoth, Mike Jr</i>			Date: <b>5/15/19</b>		Time: <b>3:55pm</b>		
Comments/Special Instructions:							

EMSL Analytical, Inc.'s Laboratory Terms and Conditions are incorporated into this chain of custody by reference in their entirety. Submission of samples to EMSL Analytical, Inc. constitutes acceptance and acknowledgment of all terms and conditions by Customer.





**Appendix B:  
Instrument Calibration Records**

# Certificate of Calibration

() Buck™ BioAire Pump Calibration Rotameter

( ) Buck™ BioSlide Pump Calibration Rotameter

Serial number: R14057

Date Calibrated: 1/22/19

Calibration Due Date: 1/22/20

## Flow Calibration

This is to certify that the rotameter listed above has been calibrated using a Buck Primary calibrator listed below which is calibrated according to A.P. Buck, Inc. calibration procedure APB-1, Ver. 6.2 and is traceable to the National Institute of Standards & Technology (N.I.S.T). A.P. Buck guarantees the accuracy of the rotameter to be within  $\pm 5\%$  of the actual flow rate.

AMBIENT CONDITIONS: Temperature  $74 \pm 3^{\circ}$  F Relative Humidity  $50 \pm 10\%$

Description	MFR.	Model	Serial #
Primary Calibrator	A.P. Buck Inc.	M30B	<input type="checkbox"/> A40020 <input checked="" type="checkbox"/> A40021

QA Approval By: 

Information contained in this document should not be reproduced in any form without the written consent of A.P. Buck, Inc. It is for reference only and cannot be used as a form of endorsement by any private or governmental regulatory body.

A.P. BUCK, INC.  
7101 Presidents Drive, Suite 110  
Orlando, FL 32809  
Phone: 407-851-8602  
Fax: 407-851-8910

**BUCK**  
A.P. BUCK, INC.

















# CERTIFICATE OF CALIBRATION AND TESTING

TSI Incorporated, 500 Cardigan Road, Shoreview, MN 55126 USA  
 Tel: 1-800-874-2811 1-651-490-2811 Fax: 1-651-490-3824 <http://www.tsi.com>

ENVIRONMENT CONDITIONS				<b>MODEL</b>	<b>982</b>
TEMPERATURE	75.9 (24.4)	°F (°C)		<b>SERIAL NUMBER</b>	<b>P17100006</b>
RELATIVE HUMIDITY	46	%RH			
BAROMETRIC PRESSURE	28.81 (975.6)	inHg (hPa)			

<input checked="" type="checkbox"/> AS LEFT	<input checked="" type="checkbox"/> IN TOLERANCE
<input type="checkbox"/> AS FOUND	<input type="checkbox"/> OUT OF TOLERANCE

## - CALIBRATION VERIFICATION RESULTS -

TEMPERATURE VERIFICATION				SYSTEM T-101			Unit: °F (°C)
#	STANDARD	MEASURED	ALLOWABLE RANGE	#	STANDARD	MEASURED	ALLOWABLE RANGE
1	32.0 (0.0)	32.4 (0.2)	31.0-33.0 (-0.6-0.6)	2	140.0 (60.0)	140.8 (60.4)	139.0-141.0 (59.4-60.6)

HUMIDITY VERIFICATION				SYSTEM H-102			Unit: %RH
#	STANDARD	MEASURED	ALLOWABLE RANGE	#	STANDARD	MEASURED	ALLOWABLE RANGE
1	10.0	9.6	7.8-12.2	4	70.0	69.7	67.8-72.2
2	30.0	29.7	27.8-32.2	5	90.0	89.3	87.8-92.2
3	50.0	49.9	47.8-52.2				

CO2 GAS VERIFICATION				SYSTEM G-101			Unit: ppm
#	STANDARD	MEASURED	ALLOWABLE RANGE	#	STANDARD	MEASURED	ALLOWABLE RANGE
1	0	0	0-50	4	3031	3043	2940-3122
2	518	510	468-568	5	5000	4988	4850-5150
3	1020	1030	970-1070				

CO GAS VERIFICATION				SYSTEM G-101			Unit: ppm
#	STANDARD	MEASURED	ALLOWABLE RANGE	#	STANDARD	MEASURED	ALLOWABLE RANGE
1	36	36	33-39	2	101	100	98-104

*TSI does hereby certify that the above described instrument conforms to the original manufacturer's specification (not applicable to As Found data) and has been calibrated using standards whose accuracies are traceable to the United States National Institute of Standards and Technology (NIST) or has been verified with respect to instrumentation whose accuracy is traceable to NIST, or is derived from accepted values of physical constants. TSI's calibration system is registered to ISO-9001:2015.*

Measurement Variable	System ID	Last Cal.	Cal. Due	Measurement Variable	System ID	Last Cal.	Cal. Due
Temperature	E003986	02-14-18	08-31-18	Temperature	E003987	02-14-18	08-31-18
Humidity	E003539	02-22-18	08-31-18	5000 CO2	c5732043	04-16-18	10-04-20
200 CO	CC506122	01-24-18	01-25-26	N2	t78516	04-17-18	04-03-23
Air	108551y	04-23-18	03-09-20	Flow	E003298	10-25-17	10-31-18
Flow	E004631	10-25-17	10-31-18	Flow	E003980	03-28-18	03-31-19
Flow	E003525	01-10-18	01-31-19	2000 C4H8	EB0053919	10-20-17	10-20-21
100 C4H8	EB0078607	09-28-16	09-28-20				

Chimona

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CALIBRATED

May 29, 2018

DATE